

## Synthesis and Pharmacological Activities of 13-Dehydro Derivatives of Primary Prostaglandins

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### Abstract

13-Dehydro derivatives of prostaglandin E<sub>1</sub>, E<sub>2</sub>, E<sub>3</sub>, F<sub>1α</sub> and F<sub>2α</sub> were synthesized. Compared with natural prostaglandins, 13-dehydro analogues were found to exhibit more potent inhibitory activity against human platelet aggregation and relaxation of guinea-pig isolated trachea, while they showed less potent activity of contraction of guinea-pig isolated ileum. © 1998 Elsevier Science Ltd. All rights reserved.

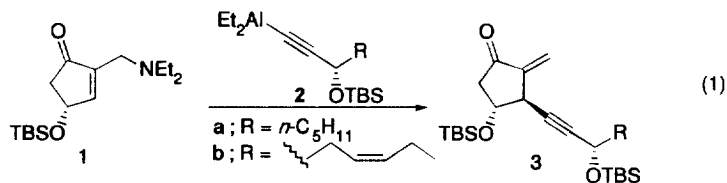
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The synthesis and biological effects of 13-dehydro derivatives of natural prostaglandins (PGs) have attracted much interest. Fried *et al.* [1,2] reported the synthesis of 13-dehydro derivatives of natural PGE<sub>2</sub> and F<sub>2α</sub>. These derivatives caused stimulation of cAMP synthesis in mouse ovary, and also caused termination of pregnancy in hamsters. Furthermore, they have proved to be nonsubstrates for 15-dehydrogenase [1,3]. PG analogues in which the double bond at C-13 has been replaced by a triple bond have been developed, and some of these have deserved particular attention as promising therapeutic agents [4–13].

13-Dehydro derivatives of natural PGE<sub>1</sub>, F<sub>1α</sub> and E<sub>3</sub>, however, have not been synthesized yet, in spite of the great interest in their biological effects. Herein we report a highly efficient synthesis and biological evaluation of 13-dehydro PGs including 13-dehydro PGE<sub>1</sub>, F<sub>1α</sub> and E<sub>3</sub>.

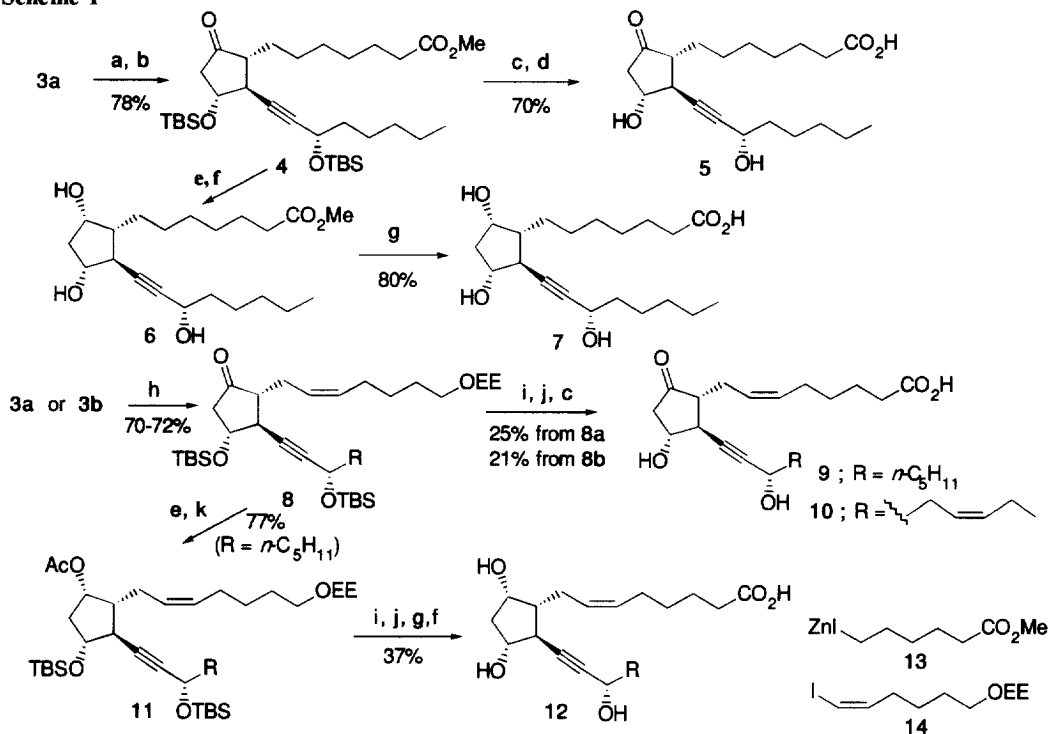
## 1. Synthesis

In a recent paper, we have reported the synthesis of  $\alpha$ -methylenecyclopentanone **3**[14], which is a potential intermediate of the synthesis of 13-dehydro PGs *via* 1,4-addition reaction of  $\alpha$  side-chain units, by the reaction of commercially available cyclopentenone **1** with alkynylaluminium compounds **2**(eq 1).



By starting from **3**, a variety of 13-dehydro PGs have been synthesized according to the procedure shown in Scheme 1. Thus, the compound **3a** reacted with an organocopper reagent derived from **13**(2.0 eq.) and CuCN-2LiCl(2.5 eq.) in the presence of trimethylsilyl chloride (1.8 eq.) to afford **4** in 78% yield[15,16].

Scheme 1



(a) **13**, CuCN-2LiCl, Me<sub>3</sub>SiCl, THF, -78°C; (b) 1N-HCl, MeOH, THF, 0°C; (c) (HF)<sub>n</sub>-pyridine, CH<sub>3</sub>CN, 0°C; (d) porcine liver esterase, phosphate buffer(pH=8.0), room temperature; (e) L-selectride, THF, -78°C; (f) aqueous HF, THF, 0°C; (g) LiOH, MeOH or EtOH, H<sub>2</sub>O then 1N-HCl, room temperature; (h) **14**, *t*-BuLi, Et<sub>2</sub>O then (2-thienyl)Cu(CN)Li, THF, -78°C to 0°C (i) pyridinium *p*-toluenesulfonate, Et<sub>2</sub>O, *i*-PrOH, room temperature; (j) Jones' reagent, acetone, Et<sub>2</sub>O, 0°C; (k) Ac<sub>2</sub>O, pyridine, cat. N,N-dimethylaminopyridine, room temperature

Protodesilylation of **4** with  $(\text{HF})_n$ -pyridine followed by hydrolysis using porcine liver esterase provided 13-dehydro PGE<sub>1</sub> (**5**)<sup>1</sup> in 70% yield. Meanwhile, 13-dehydro PGF<sub>1</sub> (**7**)<sup>2</sup> was prepared by the stereospecific reduction of a carbonyl group in **4** followed by protodesilylation and saponification. Similarly, 13-dehydro PGE<sub>2</sub> (**9**) ( $[\alpha]_{\text{D}}^{23}$  -15.4° (c 0.052, EtOH); lit.  $[\alpha]_{\text{D}}^{20}$  -15.1° (c 0.10, EtOH)) and -PGF<sub>2 $\alpha$</sub>  (**12**) ( $[\alpha]_{\text{D}}^{23}$  +36.4° (c 0.188, EtOH); lit.  $[\alpha]_{\text{D}}$  +34.0° (c 0.66, EtOH)) were synthesized from **3a** and an organocopper reagent derived from **14** [18] via the compound **8a** in 18% and 21% overall yields, respectively. Similar synthetic reactions starting from **3b** and **14** provided 13-dehydro PGE<sub>3</sub> (**10**)<sup>3</sup> via the compound **8b** in 15% overall yield.

## 2. Biological activity

The results of pharmacological evaluation of these analogues are summarized in Table 1. The biological activities of all five 13-dehydro derivatives were compared with those of the corresponding natural compounds. 13-Dehydro PGE<sub>1</sub> (**5**) showed 3.0 times more potent inhibitory effect on adenosine diphosphate (ADP)-induced human platelet aggregation *in vitro* [19] in comparison with natural PGE<sub>1</sub> and it was 8.9 times more potent against histamine-induced relaxation of guinea-pig isolated trachea precontracted with histamine [20]. On the other hand, it proved to be less potent on contraction of guinea-pig isolated ileum [21]. 13-Dehydro PGE<sub>2</sub> (**9**) and PGE<sub>3</sub> (**10**) also showed the same tendency of more potent activities of anti-aggregation and relaxation of trachea, and less potent activity on contraction of ileum. 13-Dehydro PGF<sub>1 $\alpha$</sub>  (**7**) and PGF<sub>2 $\alpha$</sub>  (**12**) proved to be 0.02 to 0.12-fold less potent than the corresponding natural compounds on contraction of guinea-pig ileum, and they showed almost the same potency on contraction of rat isolated uterus [22]. It is a noteworthy fact that the activity profiles of prostaglandins can be greatly altered by the structural modification of the 13,14-double bond to a triple bond. Of prime interest is the highly significant dissociation in 13-dehydro PGE<sub>1</sub> (**5**) of the inhibitory activity against human platelet aggregation from the contraction activity of guinea-pig ileum which is an *in vitro* system that often predicts “diarrhea” potential, thus 13-dehydro PGE<sub>1</sub> (**5**) may be a more selective anti-aggregating agent than natural PGE<sub>1</sub>.

- <sup>1</sup> <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300MHz)  $\delta$  ppm: 0.89(t, J=6.3Hz, 3H), 1.14–1.91(m, 18H), 2.10–2.46(m, 1H), 2.24(dd, J=18.2, 9.6Hz, 1H), 2.33(t, J=7.1Hz, 2H), 2.52–2.86(m, 1H), 2.75(dd, J=18.2, 7.1Hz, 1H), 4.18–4.49(m, 1H), 4.39(t, J=6.3Hz, 1H); IR(KBr): 3839, 2933, 2860, 2237, 2217, 1741, 1731, 1713, 1462, 1409, 1234, 1078, 727cm<sup>-1</sup>; MS(FAB)(+KI) m/z: 391(MK<sup>+</sup>); HRMS(FAB) m/z; Calcd for C<sub>20</sub>H<sub>32</sub>O<sub>5</sub>: 353.2328(MH<sup>+</sup>). Found: 353.2317.; mp 48.8–50.4°C (colorless needles, recrystallized from AcOEt-hexane); Anal Calcd for C<sub>20</sub>H<sub>32</sub>O<sub>5</sub> · 1/2H<sub>2</sub>O: C, 66.45; H, 9.20. Found: C, 66.35; H, 9.24.;  $[\alpha]_{\text{D}}^{27}$  -35.05° (c 0.884, MeOH)
- <sup>2</sup> <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300MHz)  $\delta$  ppm: 0.89(t, J=6.9Hz, 3H), 1.24–1.87(m, 20H), 2.13–2.24(m, 1H), 2.35(t, J=7.1Hz, 2H), 2.54–2.61(m, 1H), 4.17–4.27(m, 2H), 4.37(dt, J=6.6, 1.9Hz, 1H); IR(KBr): 3460, 3339, 2955, 2929, 2858, 2232, 1720, 1672, 1620, 1469, 1404, 1330, 1283, 1230, 1186, 1155, 1134, 1060, 1040, 1026, 978, 937, 888, 802, 727, 653, 522cm<sup>-1</sup>; MS(FAB)(+KI) m/z: 393(MK<sup>+</sup>); HRMS(FAB) m/z; Calcd for C<sub>20</sub>H<sub>34</sub>O<sub>5</sub>K: 393.2043(MK<sup>+</sup>). Found: 393.2030.
- <sup>3</sup> <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300MHz)  $\delta$  ppm: 0.98(t, J=7.5Hz, 3H), 1.64–1.78(m, 2H), 2.00–2.25(m, 5H), 2.29–2.53(m, 7H), 2.61–2.82(m, 2H), 4.28–4.49(m, 2H), 5.30–5.67(m, 4H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75MHz)  $\delta$  ppm: 14.2, 20.8, 24.6, 25.2, 26.3, 33.2, 35.6, 41.1, 46.0, 55.3, 62.2, 72.8, 84.0, 84.4, 122.8, 125.9, 132.0, 135.6, 178.0, 213.2; IR(neat): 3417, 2929, 2220, 1730, 1407, 1247, 1047, 866, 756cm<sup>-1</sup>; MS(FAB)(+KI) m/z: 387(MK<sup>+</sup>); HRMS(FAB) m/z; Calcd for C<sub>20</sub>H<sub>28</sub>O<sub>5</sub>K: 387.1574(MK<sup>+</sup>). Found: 387.1581.

**Table 1**

Relative potency of 13-dehydro prostaglandins (in vitro assay)

Compound	Platelet <sup>a,b,c</sup>	Trachea <sup>b,d,e</sup>	Ileum <sup>a,h,i</sup>	Uterine <sup>h,i,m</sup>
5	3.0 × PGE <sub>1</sub>	8.9 × PGE <sub>1</sub>	0.23 × PGE <sub>1</sub>	not tested
9	3.4 × PGE <sub>2</sub>	2.6 × PGE <sub>2</sub> <sup>f</sup>	0.18 × PGE <sub>2</sub> <sup>j</sup>	not tested
10	2.6 × PGE <sub>3</sub>	5.4 × PGE <sub>3</sub>	0.26 × PGE <sub>3</sub>	not tested
7	not tested	not tested	0.02 × PGF <sub>1α</sub>	1.00 × PGF <sub>1α</sub>
12	not tested	not tested	0.12 × PGF <sub>2α</sub> <sup>k</sup>	0.60 × PGF <sub>2α</sub> <sup>k</sup>

<sup>a</sup>Inhibition of ADP-induced human platelet aggregation.<sup>b</sup>The activities relative to those of corresponding natural PGs were calculated based on IC<sub>50</sub> values (N=4).<sup>c</sup>IC<sub>50</sub> values of natural PGE<sub>1</sub>=77.6nM, PGE<sub>2</sub>=0.23μM, PGE<sub>3</sub>=10.0μM.<sup>d</sup>Relaxation of guinea-pig trachea precontracted with histamine.<sup>e</sup>IC<sub>50</sub> values of natural PGE<sub>1</sub>=0.16μM, PGE<sub>2</sub>=18.2nM, PGE<sub>3</sub>=0.48μM.<sup>f</sup>13-Dehydro PGE<sub>2</sub>-induced relaxation of guinea-pig trachea precontracted with carbachol was reported [23].<sup>g</sup>Contraction of guinea-pig ileum.<sup>h</sup>The activities relative to those of corresponding natural PGs were calculated based on ED<sub>50</sub> values (N=4).<sup>i</sup>ED<sub>50</sub> values of natural PGE<sub>1</sub>=47.9nM, PGE<sub>2</sub>=22.9nM, PGE<sub>3</sub>=0.12μM, PGF<sub>1α</sub>=0.12μM, PGF<sub>2α</sub>=47.9nM.<sup>j</sup>Similar result was reported [21].<sup>k</sup>Similar result was reported [24].<sup>l</sup>Contraction of rat isolated uterine.<sup>m</sup>ED<sub>50</sub> values of natural PGF<sub>1α</sub>=0.17μM, PGF<sub>2α</sub>=44.7nM.

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